

## تأثير إضافة (بذور الحلبة) على صفات السائل المنوي في ذكور الأرانب جمال المختار مبارك - قسم علم الحيوان - كلية العلوم - جامعة صبراتة

### الملخص العربي

أجريت هذه الدراسة بهدف تقييم تأثير إضافة مستويات مختلفة من بذور الحلبة إلى علائق ذكور الأرانب البالغة في صفات السائل المنوي، استخدمت 32 ذكر من الأرانب البالغة (بعمر 8 أشهر) خلال الفترة من ديسمبر إلى فبراير، وقسمت إلى أربعة مجاميع وبواقع 8 أرانب لكل معاملة والمعاملات كانت كالآتي: C = معاملة السيطرة غذيت على عليقة قياسية (بدون اي اضافة) ، T1 = غذيت على عليقة السيطرة + 5 جرام / كجم علف من مسحوق بذور الحلبة، T2 = غذيت على عليقة السيطرة + 10 جرام / كجم علف من مسحوق بذور الحلبة ، T3 = غذيت على عليقة السيطرة + 15 جرام / كجم علف من مسحوق بذور الحلبة. تم جمع عينات السائل المنوي بعد مرور أربعة أسابيع من المعاملة بواسطة المهيل الأسطناعي لدراسة صفات السائل المنوي من حيث حجم القذفة والحركة الفردية والجماعية للحيوانات المنوية وحيوية الحيوانات المنوية وقيم pH ، PSV ، تركيز الحيوانات المنوية، العدد الأجمالي للحيوانات المنوية لكل قذفة إضافة إلى صفات بلازما السائل المنوي والمتمثلة في مستوى الكوليستيرول، البروتين الدهني العالي الكثافة، البروتين الدهني المنخفض الكثافة، الليبيدات الكلية، الجليسيريدات الثلاثية، ومستوى الفركتوز، TAC ، MDA ، ونشاط انزيمات AST ، ALT ، ALP .

أظهرت النتائج انخفاض معنوي للمعاملات المحتوية على بذور الحلبة في كل من حجم القذفة والحركة الفردية والجماعية وحيوية الحيوانات المنوية و PSV وتركيز الحيوانات المنوية والعدد الكلي للحيوانات المنوية لكل قذفة وانخفاض في مستوى الكوليستيرول و LDL ، والليبيدات الكلية والجليسيريدات الثلاثية وانزيمات ALP ، AST ، MDA مقارنة بمجموعة السيطرة. وزيادة معنوية في الحيوانات المنوية الميتة و LDL ، TAC مقارنة بمجموعة السيطرة في حين لا توجد اختلافات في قيم pH مابين مجموعة السيطرة والمعاملات المحتوية على بذور الحلبة. نستنتج من هذه الدراسة أن إضافة مستويات مختلفة من بذور الحلبة وخاصة مستوى 15 جرام/ كجم يؤدي إلى انخفاض معنوي في صفات السائل المنوي لذكور الأرانب.

### EFFECT OF NATURAL ADDITIVE (FENUGREEKSEEDS) ON SEMEN TRAITS IN BUCKS

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**ABSTRACT** This study was conducted to evaluate the effect of the addition of fenugreek seeds (F.S) to the rabbit bucks ration on semen quality and semen plasma traits. A total number of 32 male New Zealand rabbits (8 month old) divided into four groups (8 males/group) and treated as follows T1 = control group reared on standard ration, T2 = standard ration supplemented with 5gm F.S /kg, T3 = standard ration supplemented with 10gm F.S /kg, T4 = standard ration supplemented with 15gm F.S /kg. Semen samples were assessed after four weeks of treatment as regards ejaculation volume, mass activity, Individual motility, pH, PSV, semen concentration and totalsperm count. However, the second semen collection was used for determine seminal plasma concentrations of cholesterol, LDL, HDL, total lipids, triglycerides, fructose level, MDA, TAC, and activities of aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) enzymes. Results revealed that the treatment (T2, T3 and T4) showed that a significant ( $p \leq 0.05$ ) decrease in ejaculation volume, mass activity, individual motility, PSV, semen

concentration and total sperm count and seminal plasma cholesterol, LDL, total lipids, triglycerides, ALP, AST, ALT enzyme and MDA, and significantly ( $P < 0.05$ ) increase in dead sperm and seminal plasma concentrations of HDL, TAC as compared with the control group. There was no significant difference in pH of semen among the control and experimental groups (T1, T2 and T3). In conclusion, the results of this study showed that supplementing fenugreek seeds in the diet of buck rabbits resulted in significant decrease in semen quality and seminal plasma characteristics.

**Keywords:** Fenugreek seeds, Rabbit bucks, semen quality.

## INTRODUCTION

Feed additive are important materials that can improve the efficiency of feed utilization and animal performance. The possibility of using new natural alternative additives instead of antibiotics and hormone in animal diets is being recently used. Some plants, containing various essential oils, have been used as alternative remedies

by some researchers (Ceylan *et al.*, 2003). Fenugreek (*Trigonella foenum graecum* L.) is an annual plant from the family of Leguminosae, originate in India and native cultivated in south Europe, Northern Africa, India and Egypt and south of Iraq. It has many uses in medical purposes, fenugreek seeds have a great effect in relation to increasing lactation; also it helps to increase urine and menstruation in women. It helps also in rickets and anemia. Owing to the existence of mucilage, it helps relieve sore throat and is useful in the treatment of asthma and difficult breathing. Fenugreek is considered as an appetizer and helps in digestion. Fenugreek seeds have also been recognized as a potential source of diosgenin, a basic compound in the hemisynthesis of steroidal sapogenins such as cortisone and sex hormones (Brenac and Sauvaire 1996a,b). It is used as astringent materials and anti-bloat (Chopra *et al.*, 1982), and used in animal feed (Cheij, 1984). However it is used as a supplement to poultry feeding to lower plasma total lipids and total cholesterol in Hubbard broiler chicks (Azouz, 2001) and improve antioxidant status and production performance in laying hens (Alkatan, 2006). Fenugreek seeds improve the reproductive and physiological performance of broiler breeder males (Taha, 2008), and revealed positive significant results of semen traits in aged broiler breeder males (Abdul-Rahman *et al.* 2010).

Fenugreek like other legumes is a good source of dietary protein (20-30%) for consumption by human and animals, the fatty acids from 5-10% which are predominantly linoleic, linolenic, oleic and palmitic acids. It has 45-65% total carbohydrates with 15% of galactomannan (Schryver, 2002). Fenugreek seeds have been used extensively to prepare extract and powder for medicinal uses (Basch *et al.*, 2003). It is reported to have anti-diabetic, anti-fertility, anti-cancer, anti-microbial, anti-parasitic, hypo-cholesterolaemic effects (Al-Habori and Roman, 2002). Also, it contains many minerals and vitamins (Micheal and Kumawat, 2003). (Seleem, 2008) found that supplementation of 0.3% fenugreek to rabbit diet showed a great role in enhancing the immune systems, improved growth performance, blood metabolites and reproductive performance. A study showed that in rabbits, a fenugreek seed powder containing diet (30%) for 3 months could significantly decrease male testis weight [ $\sim 25\%$ ] and sperm concentration [ $\sim 43\%$ ], indicating a toxic effect of fenugreek seeds on seminiferous tubules and the interstitial tissue (Leydig cells). The negative impact of fenugreek seeds on the male structural and functional integrity of testicular tissues was evidenced by the histopathological data highlighting the damage of interstitial tissue, showing a decrease in the number of seminiferous tubules with mild spermatogenesis hypoplasia when compared with that in the control group (Kassem *et al.*, 2006). Therefore the present study was designed to examine the ability of fenugreek seeds to decrease in semen quality and seminal plasma characteristics in the rabbit bucks.

## MATERIALS AND METHODS

The present study was carried out at the Rabbit Research Laboratory, Department of Animal Production, Faculty of Veterinary Science, Agriculture, Zawia University during the period from December 2016 to February 2017, to determine the effect of addition of fenugreek seeds to the rabbit bucks ration on semen quality and the seminal plasma traits.

A total of 32 males of New Zealand rabbit bucks 8 months old of proven fertility, with an average initial live body weight of  $2.85 \pm 0.03$  kg, were randomly distributed into were allocated for the 4 treatment groups containing 8 bucks each. Treatment groups were as follows: Control diet (free from fenugreek seeds), T1: Control diet + 5 g/kg fenugreek seeds, T2: Control diet + 10g/kg fenugreek seeds; T3: Control diet + 15 g/kg fenugreek seeds. Bucks were allowed to become accustomed to treatment for a preliminary period of 28day during December 2016. Bucks were housed individually in galvanized batteries (50Lx50Wx40Hcm) provided with feeders and automatic drinkers in a windowed rabbitary.

Semen samples were collected weekly over 8 weeks using an artificial vagina and the samples of the each week were subjected to chemical analysis. Semen collection and handling were carried out and evaluated according to the international guidelines of (IRRG, 2005). Ejaculated volume was measured to the nearest 0.01 ml. A weak eosin- formalin (10% formalin) solution was used for evaluation of sperm concentration by the improved Neubauer hemocytometer slide method as described by **Smith and Mayer (1995)**. Total sperm out- put was calculated by multiplying semen ejaculation volume by semen concentration. Semen mass motility was given an arbitrary score from 0 to 3 based on the following assessment and the following variables were estimated: 0=No current, (0.5) = Very few slow current, 1 = Few slow current, 1.5= Many moderate waves, 2= Many sweeping waves, 2.5= numerous vigorous waves, 3= numerous rapid and vigorous waves, as described by **Moule (1995)**. Individual sperm motility was estimated at 400x magnification (**Kamar, 1960**). Evaluation of seminal initial fructose was carried out immediately after collection according to **Mann (1948)**. Assessments of dead and abnormal spermatozoa were performed using an eosin-aniline blue staining mixture (**Shaffer and Almquist, 1948**). Initial hydrogen ion concentration (pH) of semen sample was determined just after collection using pH meter. Packed sperm volume (PSV) was recorded using Micro-AID® microhematocrit tubes and microhematocrit-centrifuge which centrifuged for 5 min at 4000 RPM. Seminal plasma was separated by centrifugation at 3000 rpm for 20 minutes and was stored at -20°C in Eppendorf tubes for further analysis of total lipids, cholesterol, LDL, HDL, total antioxidants capacity, malondialdehyde, triglycerides, alkaline phosphatase, ALT and AST which were determined in seminal plasma colorimetrically using commercial kits obtained from (BIO- DIAGNOSTICS, Egypt) according to the procedure outlined by the manufacturer.

### Statistical analysis

Results are expressed as mean  $\pm$  standard error (SE). Differences between means in different group were tested for significance using a one- way analysis of variance (ANOVA) followed by Duncan's multiple range test (**Duncan, 1955**). The *P* value of 0.05 or less was considered significant using **SPSS (2004)**. The model was:

$Y_{ij} = \mu + T_i + e_{ij}$  where  $Y_{ij}$  = an observation treatment;  $\mu$  = the overall mean;  $T_i$  = effect of treatments groups and  $e_{ij}$  = the random error component assumed to be normally distributed.

## RESULTS AND DISCUSSION

Fenugreek seeds supplementation caused a significant decrease ( $p \leq 0.05$ ) in ejaculation volume, mass and individual motility, spermatozoa concentration and total sperm count in comparison with the control group, and a significant increase in dead sperm percentage as compared with the control group, while there was no significant differences among treatment groups in relation to the pH of

semen (**Table 1**). Including fenugreek seeds in the diet resulted in significant ( $P \leq 0.01$ ) decrease in ejaculate volume, mass activity, individual sperm motility, PSV%, sperm concentration and total sperm count in comparison with the control group. While dead sperm percentage was numerically increased in comparison with the control group. **Kassem et al., (2006)** found that feeding diets containing 30% fenugreek seeds to male and female white New Zealand rabbits clearly demonstrate an antifertility effect of fenugreek seeds in the female rabbits and more of a toxicity effect in the male rabbits. The results showed that diets containing 30% fenugreek seeds significantly reduced male testis weight (~25%) as well as sperm concentration (~ 43%), indicating a toxic effect of fenugreek seeds on seminiferous tubules and the interstitial tissue (Leydig cells). Furthermore, feeding fenugreek seeds at 30% to male rabbits lowered circulating androgen (testosterone) by 65.8%. Showed that the negative impact of fenugreek seeds on the male structural and functional integrity of testicular tissues was evidenced by the histopathological data highlighting the damage of interstitial tissue, showing a decrease in the number of seminiferous tubules with mild spermatogenesis hypoplasia when compared with that in the control animals. The latter finding may support the hypothesis that a component of fenugreek seeds might have a direct toxic effect on the cells responsible for synthesis of androgens (**Kamal et al., 1993**). This is further supported in the report of **Kassem et al., (2006)**, by the lack of differences in the number of litter size when treated males were mated with control females, despite the decrease in androgen levels and sperm concentration, suggesting a toxicity effect rather than an antifertility effect in the male rabbits. **AlYahya, (2013)** reported that oral administration of fenugreek at the higher doses of 305 and 610 mg/kg body weight per day has caused male reproductive toxicity followed by reduced fertility, decreased motility, sperms count and an increase in the proportion of abnormal sperms in mice. The reduction observed in the fertility of male mice and the abnormal shapes of the sperms observed might be related with the increased accumulation of free radicals. Previous studies (**Farag et al., 2010; Al-Majed et al., 2006**) also showed that the depletion of glutathione cause spermatotoxicity and produce abnormal shapes of the sperms. Furthermore, the endocrinological study in plasma of male mice showed increase of estradiole and reduction of testosterone, which are known to affect the fertility. These results confirms the previous reports (**Kassem et al., 2006; Khare et al., 1983**) which showed fenugreek seeds to demonstrate an anti-fertility effect in both male and female rabbits. **Khalki et al., (2010)** reported that aqueous seeds extract of *Trigonella foenum-graecum* affected reproduction in mice and showed teratogenic and foetotoxic effect. Steroidal saponins and alkaloids have been shown to be teratogenic. **Araee et al., (2009)** has considered that in view of the presence of the steroidal saponin diosgenin in fenugreek seeds, it is likely that the administration of fenugreek in high doses adversely influences bone marrow cell proliferation. These changes are attributed to the toxic constituents present in fenugreek, such as quinones and flavonoids (**Skibola and Smith, 2000; Wu et al., 1997**). **Mohammed et al., (2013)** reported that *Trigonella foenum-graecum* seeds ethanolic extract decreased serum testosterone concentration in the treated rats and both mass and individual motility of the sperms in the treated cocks, while it insignificantly increased abnormal sperm percentage. Histopathological examination of rat's treated with *Trigonella foenum-graecum* seeds ethanolic extract showed presence of inactive seminiferous tubules and oedema formation between the seminiferous tubules. This effect may also contribute to the potential antifertility effects of this extract. The primary function of testosterone is the maintenance of spermatogenesis and hence fertility also testosterone is important in the maturation of spermatozoa as they pass through the epididymis and vas deferens. A low testosterone level is one of the best indicators of hypogonadism of hypothalamic or pituitary origin, although very high levels of testosterone suppress sperm autogenesis (**McLachlan et al., 2002**). **Jankorvaa et al., (2003)** had reported that 12.6% of male infertile patients have hypotestosteronaemia. The decrease of sperm concentration in cocks may be due to the fact that *Trigonella foenum-graecum* increases the prolactin levels and higher levels of prolactin tends to inhibit production of GnRH leading to decrease in both FSH and LH and thereby decrease spermatogenesis and effects, in male, that mentioned by previous study of **Kassem et al., (2006)** which reported that high concentrations of fenugreek in the diet significantly reduces fertility of

both male and female rabbits and reduced testis weight, with evident damage to the seminiferous tubules and interstitial tissues. **Embark (2016)** found that the supplementation of male rabbit's diets with 6g/kg fenugreek seeds decreased reproductive performance.

The effect of fenugreek seeds on seminal plasma total lipids, triglycerides, total cholesterol, high density lipoprotein (HDL), low density lipoprotein (LDL), AST, ALT, ALP, fructose level, TAC, and MDA of bucks are presented in **Table (2)**.

Seminal plasma total lipids, triglycerides and cholesterol were significant ( $P \leq 0.01$ ) decreased in rabbits fed diets containing fenugreek seeds. Fenugreek seed decreased seminal plasma total lipids, triglycerides and cholesterol in comparison with the control group. Fenugreek seeds used in the present study as phytobiotics resulted in decreasing seminal plasma LDL levels in comparison with the control group, while seminal plasma HDL was significantly increased in these treatments when compared with the control group. **Tavilani et al., (2014)** studied the correlation between serum lipid concentrations and variations in seminal lipid parameters in infertile men. They found no relationship between the concentration of cholesterol, phospholipids and triacylglycerols in serum, spermatozoa or seminal plasma of the infertile men, which is consistent with the findings of several other authors (**Grizard et al., 1995**). **Grizard et al., (1995)** compared the effect of hypercholesterolemia and normocholesterolemia on the spermatozoa and seminal content of cholesterol and phospholipids. They suggested that hypercholesterolemia has no effect on cholesterol and phospholipid levels in spermatozoa and seminal plasma (**Grizard et al., 1995**). Since cholesterol has a major role in the sperm membrane, which is essential for sperm cell function, it can be assumed that an increase of cholesterol level in the blood will also increase the cholesterol content of semen. This hypothesis was not confirmed in the study of **Tavilani et al., (2014)**. There appears to be no correlation between the amount of cholesterol in the serum and in sperm or seminal plasma, suggesting that sperm cholesterol content is regulated locally within the male reproductive tract (**Travis and Kopf, 2002; Saez et al., 2011; Maqdasy et al., 2012**). For proper function of spermatozoa, cholesterol and phospholipids should be regulated accurately. In the male reproductive tract lipid homeostasis is done by testicular and post-testicular function (**Maqdasy et al., 2012, Lobaccaro et al., 2012**). The results of **Tavilani et al., (2014)** showed no correlation between serum lipids with sperm parameters, which is consistent with the findings of some other authors. **Khalili et al. (2009)** reported that the concentrations of serum lipids were not generally related to the quality of semen parameters. Nonetheless, the results of **Tavilani et al., (2014)** were not consistent with the results of some other studies, in which animals were fed with a high-fat diet (**Shalaby et al., 2004; Saez Lancellotti et al., 2010**). These results suggest that serum cholesterol, phospholipids and triacylglycerols have no effect on the levels of cholesterol, phospholipids and triacylglycerols in spermatozoa and seminal plasma, and in addition, they do not cause any alteration of semen parameters. Seminal plasma alkaline phosphatase (ALP), ALT and AST showed significant ( $P \leq 0.01$ ) decrease in rabbits fed diets containing fenugreek seeds comparison with the control group. These results get along with the finding of (**Roussal and Stalleup, 1965**) who found negative correlation between AST activity of seminal plasma and with each of ejaculation volume, sperm motility, sperm concentration and percent live cells. Also **Chauban et al., (1993)** found a positive correlation between enzyme release and sperm acrosomal damage. The results presented in **Table (2)** showed significant increase ( $P \leq 0.01$ ) in seminal plasma fructose level and this increase in rabbits fed diets containing fenugreek seeds as compared with control group. In seminal vesicle, the seminal plasma is synthesized which is a medium for sperms. It consists of proteins, fructose, mucus, vitamin c, flavins, phosphoryl choline and prostaglandins. The high fructose concentration provides nutrient energy for the spermatozoa (**Wilke et al., 2009**), which reflects testosterone action and better quality of semen (**Taha, 2008**).

In the present study, seminal plasma total antioxidant capacity increased significantly ( $P \leq 0.01$ ) as a result of feeding rabbits on diet containing fenugreek seed as compared with the control group. In support of this finding, lipid peroxidation (malondialdehyd) levels of rabbits fed fenugreek seed in significant ( $P \leq 0.01$ ) decrease as compared with the control group. Reactive

oxygen species are metabolites of oxygen, including hydrogen peroxide and the superoxide and hydroxyl radicals (Mostafa *et al.*, 2006). The mammalian spermatozoon can generate ROS under physiologic conditions to mediate signal transduction, as well as to regulate the sperm function (Naughton *et al.*, 2001). On the other hand, like most cells, spermatozoa are equipped with antioxidant enzymes such as superoxide dismutase, glutathione peroxidase, and catalase to detoxify harmful excessive ROS and to prevent cell injury (Suleiman *et al.*, 1996). The antioxidant-rich secretions of the seminal vesicles, including both enzymatic and nonenzymatic antioxidant systems, also assist in protecting the spermatozoa from ROS (Werthman *et al.*, 2007). Reduced glutathione (GSH) is an important component of antioxidant systems. Under high stress conditions, GSH regeneration is retarded and the antioxidant capacity of the tissue is impaired. Under such circumstances, ROS production surpasses the antioxidant capacity and causes increased oxidative stress (Naughton *et al.*, 2001). In support of this account, exogenous glutathione is known to ameliorate the effects of varicocele and partially to restore fertility (Lenzi *et al.*, 1994). Owing to the low amount of cytoplasm and abundance of polyunsaturated fatty acids in the plasma membrane of spermatozoa, spermatozoa are susceptible to damage by oxidative stress through ROS, especially lipid peroxidation (Chen *et al.*, 2008; Gonzales, 2001; Zalata *et al.*, 2004). This may result in acidification by some or all of the following mechanisms. Malondialdehyde, produced by the peroxidation of polyunsaturated fatty acids, may be oxidized to malonic acid, which potentially can be decarboxylated to acetate, as in the liver (Siu and Draper, 1982; Davydov, 1993). Polyamines, particularly spermine, are essential for spermatozoal activity (Atmar *et al.*, 1981). Malondialdehyde will react with spermine and other amines, forming Schiff bases. This will further decrease the intracellular pH, as well as directly impairing spermine-dependent cellular functions. The optimum pH for ROS scavenging by the enzymatic antioxidant systems ranges between neutral and slightly alkaline (BRENDA, 1999). Thus, lowering of the pH markedly depresses the antioxidant enzyme activities, which indeed are significantly impaired in varicocele cases (Barbieri, 1999). Therefore, there is a directly proportional relationship between acidification and increasing ROS levels.

## Conclusion

Conclusion, the results of this study showed that supplementing fenugreek seeds in the diet of buck rabbits resulted in significant decreased in semen quality and seminal plasma characteristics.

**Table1: Effect of dietary supplementation with different levels of fenugreek seeds on semen traits (Mean ± SE) of buck rabbits**

Traits	Treatments				P.Value
	Control	T1	T2	T3	
Ejaculate volume(ml)	0.71±0.003 <sup>a</sup>	0.67±0.003 <sup>b</sup>	0.62±0.003 <sup>c</sup>	0.57±0.004 <sup>d</sup>	0.0001
Mass activity (1-3)	2.33±0.003 <sup>a</sup>	2.27±0.003 <sup>b</sup>	2.22±0.003 <sup>c</sup>	2.17±0.004 <sup>d</sup>	0.0001
Individual motility %	77.06±0.19 <sup>a</sup>	75.05±0.22 <sup>b</sup>	73.19±0.33 <sup>c</sup>	70.87±0.35 <sup>d</sup>	0.0001
Dead sperm %	20.16±0.29 <sup>d</sup>	21.84±0.21 <sup>c</sup>	23.39±0.28 <sup>b</sup>	25.36±0.15 <sup>a</sup>	0.0001
pH	7.95±0.007	7.96±0.005	7.94±0.003	7.96±0.004	0.876
*PSV %	12.03±0.18 <sup>a</sup>	11.61±0.12 <sup>b</sup>	10.23±0.07 <sup>c</sup>	10.18±0.03 <sup>c</sup>	0.0001
Sperm concentration (10 <sup>6</sup> mm <sup>3</sup> )	263.26±1.45 <sup>a</sup>	257.52±1.22 <sup>b</sup>	254.70±1.16 <sup>c</sup>	249.86±1.23 <sup>d</sup>	0.0001
Total sperm (10 <sup>6</sup> /ejaculate)	187.89±1.62 <sup>a</sup>	174.14±1.35 <sup>b</sup>	159.82±1.12 <sup>c</sup>	144.61±1.25 <sup>d</sup>	0.0001

Each value represented the mean of 8 semen evaluations; C: Control group; T1, T2 and T3: Diet supplemented with 5, 10 and 15 g/kg of fenugreek seeds; a-d Values within rows followed by different letters differ significantly (P<0.05).

\*PSV = Packed sperm volume.

**Table2: Effect of dietary supplementation with different levels of fenugreek seeds on seminal plasma traits (Mean ± SE) of buck rabbits**

Traits	Treatments				P.Value
	Control	T1	T2	T3	
Cholesterol(mg/dl)	54.71±0.42 <sup>a</sup>	52.05±0.52 <sup>b</sup>	49.54±0.48 <sup>c</sup>	45.74±0.38 <sup>d</sup>	0.0001
LDL (mg/dl)	19.19±0.27 <sup>a</sup>	17.13±0.21 <sup>b</sup>	15.89±0.38 <sup>c</sup>	13.63±0.27 <sup>d</sup>	0.0001
HDL mg/dl)	20.240±0.44 <sup>c</sup>	22.757±0.43 <sup>b</sup>	23.83±0.32 <sup>ab</sup>	24.91±0.34 <sup>a</sup>	0.0001
Total lipids (mg/dl)	140.15±0.47 <sup>a</sup>	137.40±0.30 <sup>b</sup>	135.15±0.47 <sup>c</sup>	128.78±0.69 <sup>d</sup>	0.0001
Triglyceride (mg/dl)	132.27±0.86 <sup>a</sup>	124.97±0.52 <sup>b</sup>	121.40±0.24 <sup>c</sup>	114.97±0.48 <sup>d</sup>	0.0001
ALP (IU/L)	44.31±0.25 <sup>a</sup>	39.45±0.27 <sup>b</sup>	37.82±0.51 <sup>c</sup>	37.19±0.30 <sup>c</sup>	0.0001
AST (U/L)	25.69±0.23 <sup>a</sup>	24.14±0.20 <sup>b</sup>	24.12±0.38 <sup>b</sup>	24.21±0.30 <sup>b</sup>	0.001
ALT (U/L)	23.52±0.39 <sup>a</sup>	22.13±0.28 <sup>b</sup>	22.86±0.37 <sup>b</sup>	22.56±0.22 <sup>b</sup>	0.001
Fructose (mg/100ml)	193.86±1.79 <sup>c</sup>	209.35±3.18 <sup>b</sup>	214.08±1.16 <sup>b</sup>	220.65±1.62 <sup>a</sup>	0.0001
TAC mmol/l	0.94±0.002 <sup>d</sup>	1.05±0.01 <sup>c</sup>	1.12±0.01 <sup>b</sup>	1.18±0.01 <sup>a</sup>	0.0001
MDA nmol/ml	5.43±0.01 <sup>a</sup>	5.31±0.06 <sup>b</sup>	5.20±0.04 <sup>c</sup>	4.89±0.04 <sup>d</sup>	0.0001

Each value represented the mean of 8 semen evaluations; C: Control group; T1, T2 and T3: Diet supplemented with 5, 10 and 15g/kg of fenugreek seeds; a-d Values within rows followed by different letters differ significantly (P<0.05).

## REFERENCES

- Abdul- Rahman ,S . Y . , SultanK. H . andTahaA .T . (2010)**. Theeffect of use of fenugreek seeds on the reproductive performance ofaged broiler breeder males .J.Of Tikrit University For Agricultural Sciences , no( 2) vol . 10 . p 156-163.
- Alkatan , M . M . (2006)** . Effect using some antioxidants on productionperformance and some physiological characters in laying hens. Ph.D.Thesis. College of Agriculture and Forestry, University of Mosul. Iraq.
- Al-Habori, M. and Roman,A. (2002)**.Pharmacological properties infenugreek-The genus Trigonella. 1st Edn. by G A Petropoulos (Ed), Taylor and Francis, London and New York. 10:163–82.
- Al-Majed, A. A., Al-Yahha,A. A. Al-Bekairi,A. M Al-ShabanahO. A,and Qureshi,S. (2006)**.Reproductive, cytological and biochemical toxicity of Yohimbein male Swiss albino mice. Asian J. Andro.1 8: 469-476.
- AlYahya, A. A., (2013)**: Reproductive, cytological and biochemical toxicity offenugreek in male Swiss albino mice. African Journal of Pharmacy andpharmacology Vol 7 (29): 2072-2082.
- Araee, M., Norouzi,M., HabibiG., and Sheikhvatan,M. (2009)**.Toxicityof Trigonella foenum-graecum L. (fenugreek) in bone marrowcell proliferation in rat. Pak. J. Pharm .Sci. 22: 126–130.
- Atmar, V.J., Kuehn,G. D. and Casillas,E. R. (1981)**.A polyamine-dependent protein kinase from bovine epididymal spermatozoa. J Biol Chem 256: 8275–8278.
- Azouz, H.M.M. 2001**. Effect of hot pepper and fenugreek seeds supplementation on broiler diets. Ph.D. Thesis, Poultry Nutrition Dept. Faculty of Agriculture, Cairo University.
- Barbieri, E.R., Hidalgo,M. E. Venegas,A. Smith,R. and Lissi,E. A. (1999)**.Varicocele-associated decrease in antioxidant defenses. J Androl 20:713–717.
- Basch, E., C. Ulbricht, G. Kuo, Szapary,P. and Smith,M. (2003)**. Therapeutic applications of fenugreek. Altern Med Rev 8: 20-27.
- Brenac, P. and Y. Sauvaire 1996a**. Chemotaxonomic value of sterols and steroidal sapogenins in the genus Trigonella. Biochem. Syst. Ecol., 24: 157-164.
- Brenac, P. and Y. Sauvaire (1996b)**. Accumulation of sterols and steroidal sapogenins in developing fenugreek pods: possible biosynthesis in situ. Phytochemistry, 41: 415-422.
- Brenda, (1999)**.The Comprehensive Enzyme InformationSystem,<http://www.brenda-enzymes.org/>. Accessed 14 June 2008.
- Ceylan, N., I. Ciftci and Z. Ilhan, (2003)**. The effect of somealternative feedadditive for antibiotic growth promoters on the performance and gut microflora of broiler chicks. Turk. J. Vet. Anim. Sci., 27: 727-733.
- Duke, J.A., M.J.B. Godwin and A.R. Ottesen, 2009.
- Chauban, M.S., Kapila,R., Gandhi,K. K, Anandm,S. R. (1993)**.Acrosome damage and enzyme leakage of goat spermatozoa during dilution, cooling and freezing. Andrologia, 26, 21-26.
- Cheij,R.1984**. McDonald Encyclopedia of medical plants . McDonald and Co., (publishers ) Ltd, London, PP, : 209,309, 313.
- Chen, S. S., Huang,W. J Chang,L. S. and Wei,Y. H. (2008)**.Attenuation of oxidative stress after varicocelectomy in subfertile patients with varicocele. J Urol 179:639–642.



- Chopra, K. L. Honda and, L. D. kapur.**1982. Chopras endogenous drag of India, Academic publisher, Calcutate, New Delhi, India.P.582.
- Davydov, V. V., (1993):** Pathways of endogenous malonate formation in the rat liver. Ukr Biokhim Zh, 65:85–88.
- Duncan, D. B.,(1955).**Multiple ranges and multiple F test. Biometrics,11:1-42.
- Embark, A., (2016).**Effect of fenugreek, anise and parsley seeds as feed additives on the productive and reproductive performance of rabbits.Thesis.Faculty of Agriculture. Alexandria-University.
- Farag, I. M., Abdel-Aziz,K. B., Nada,S. A., Tawfek,N. S., FaroukT. and Darwish,H. R. (2010).**Modulation of ochratoxin-induced oxidative stress, genotoxicity and spermatotoxic alterations by *Lactobacillus rhamnosus* in male Albino mice. J. Am. Sci. 26: 1545- 1603.
- Gonzales, G. F., (2001).**Function of seminal vesicles and their role on male fertility. Asian J Androl 3:251–258.
- Grizard, G., B. Sion, P. Jouanel, P. Benoit and D. Boucher., (1995).**Cholesterol, phospholipids and markers of the function of the accessory sex glands in the semen of men with hypercholesterolaemia. *Int J Androl.* 18(3):151-156.
- IRRG, International Rabbit Reproduction Group.,(2005):** Guidelines for the handling of rabbit bucks and semen. World Rabbit Sci., 13: 71-91.
- Jan orvaa, s o era., Laato,M andPollanen,P. (2003).**Analysis of 508 infertile male patients in south-western Finland in 1980–2000: hormonal status and factors predisposing to immunological infertility. European Journal of Obstetrics and Gynecology and Reproductive Biology 111: 173–178.
- Kamal, L. R., Yadav,R. and. Sharma,J. D. (1993).**Efficacy of the steroidal fraction of fenugreek seed extract on fertility of male albino rats. *Phytother Res,* 7:134 –138.
- Kamar, G. A. R., (1960):** The influence of semen characteristics on hatching results of chicken eggs. *Poultry Science,* 39: 188-193.
- Kassem A, Al-Aghbari A, Al-Habor, M, Al-Mamary M, (2006).** Evaluation of the potential antifertility effect of fenugreek seeds in male and female rabbits. *Contraception.*73(3):301-306.
- Khalili, M. A., Zare-Zadeh,N. Hashemi,H. (2009).**Correlation between serum lipids profile with sperm parameters of infertile men with abnormal semen analysis. *Iran J Reprod Med.* 7(3):123-127.
- Khalki, L., BaM'hamed,S. Bennis,M. Chait,A. and Sokar,Z. (2010).**Evaluation of the developmental toxicity of the aqueous extract from *Trigonella foenum-graecum* (L.) in mice. *J Ethnopharmacol.* 131: 321–325
- Khare, A. K., Sharma,M. K. andBhatnagar,V. M. (1983).**Mild anti-fertility effect of ethereal extract of seeds of *Trigonella foenum graecum* (Methi) in rats. *Arogya J Health Sci.* ;9 :91 – 93.
- Lenzi, A., M. Picardo, L. Gandini, F. Lombardo, O. Terminali and S. Passi, (1994).**Glutathione treatment of dyspermia: effect on the lipoperoxidation process. *Hum Reprod* 9:2044–2050.
- Lobaccaro, J. M. A., F. Brugnol, D. H. Volle and S. Baron, (2012).**Lipid metabolism and infertility: is there a link? *Clin Lipidol.*7(5):485-488.
- Mann, T., (1948).**fructose content and fructolysis in semen. Practical application in the evaluation of semen quality. *Journal of Agriculture Science,* 38: 323-331.
- Maqdasy, S., M. Baptissart, A. Vega , S. Baron, J. M. Lobaccaro and D. H. Volle, (2012).**Cholesterol and male fertility: what about orphans and adopted? *Mol CellEndocrinol.*368(1-2):30-46.

**McLachlan, R. I., L. O'Donnell, S. J. Meachem, P. G. Stanton, D. M. e Kretser, K. Pratis and D. M. Robertson, (2002).** Identification of specific sites of hormonal regulation in spermatogenesis in rats, monkeys, and man. *Recent Prog Horm Res.* 57:149–179.

**Michael, D. and D. Kumawat, (2003).** Legend and archeology of fenugreek, constitutions and modern applications of fenugreek seeds. *International Symp. USA,* :p. 4142.

**Mohammed, M. M., A. Mudathir , Shaddad, Sania A. I., B. Elsharif and Abu Algasem, Afaf E., (2013).** Antifertility Effects of *Trigonella foenum-graecum* (fenugreek) ethanolic extract in male rats & cocks. *J Phar m Biomed Sci.* 32: 1299-1304.

**Mostafa, T., T. H. Anis, S. Ghazi, A. R. El-Nashar, H. Imam and I. A. Osman, (2006).** Reactive oxygen species and antioxidants relationship in the internal spermatic vein blood of infertile men with varicocele. *Asian JAndrol* 8: 451–454. **Moule, G. R., (1995).** Filed investigations with sheep a manual of techniques. The common wealth scientific and industrial research organization, Australia.

**Naughton, C. K., A. K. Nangia and A. Agarwal, (2001).** Pathophysiology of varicoceles in male infertility. *Hum Reprod Update* 7:473–481.

**Roussal, J. D. and O.T. Stallcup, (1965).** Activity of lactic dehydrogenase and Its Isozymes in Bovine Semen. *J. of Dairy Science,* 48(11): 1506-1510.

**Saeid, J. M . and. AL – Soudi .K . A.(1975).** Seasonal variation in semen characteristics of White Leghorn , New Hampshire and indigenouse chicken in Iraq . *Br . Poult . Sci .* 16: 97 – 102 .

**Saez, F., Ouvrier,A. and. Drevet,J. R (2011).** Epididymis cholesterol homeostasis and sperm fertilizing ability. *Asian J Androl,* 13(1): 11-17.

**Saez Lancellotti, T. E., P. V. Boarelli, M. A. Monclus, M. E. Cabrillana, M. A. Clementi and L. S. Espinola, (2010).** Hypercholesterolemia impaired sperm functionality in rabbits. *PLoS One.* 5(10):13457.

**Schryver, T., (2002).** Fenugreek. *Total Health,* 24:42-44.

**Seleem, T.S.T., (2008).** Rabbit productivity and reproductivity as affected by fenugreek in diets. *The 1st Egyptian Conference on Rabbits Sciences:* 142-153.

**Shaffer, H. E. and Almquist,J. O. (1948).** Vital staining of bovine spermatozoa with an Asian-aniline blue staining mixture. *J. Dairy Science,* 31: 677- 678.

**Shalaby, M. A., H. Y. El-Zorba and G. M. Kamel, (2004).** Effect of alpha-tocopherol and simvastatin on male fertility in hypercholesterolemic rats. *Pharmacol Res.,* 50(2):137-142.

**Siu, G. M. and H. H. Draper, (1982).** Metabolism of malonaldehyde in vivo and in vitro. *Lipids,* 17:349–355.

**SPSS, (2004):** Statistical Package for social Sciences Release 16.0.1 Version. SPSS Inc.

**Smith, J. T. and D. T. Mayer, (1995).** evaluation of sperm concentration by the hemocytometer method. *Fertil.Steril.,* 6: 271-275.

**Suleiman, S. A., M. E. Ali, Z. M. S. Zaiu, E. M. A. El-Malik and M. A. Nasr, (1996).** Lipid peroxidation and human sperm motility: protective role of vitamin E. *J Androl* 7:530–537.

**Taha ,A.T.(2008).** The role of vitamins A , C and fenugreek seeds in lowering the effect of oxidative stress effect on physiological and reproductive performance of broiler breeder males . Ph.D. Thesis. College of Agriculture and Forestry , University of Mosul . Iraq .

**Tavilani, H ,Vatannejad .A , Akbarzadeh. M, Atabakhash .M, Khosropour. S, Mohaghgeghi .A, (2014).** Correlation between Lipid Profile of Sperm Cells and Seminal Plasma with Lipid Profile of Serum in Infertile Men. *Avicenna J Med Biochem.,* 2(1): 19607.

**Travis, A. J. and G. S. Kopf, (2002).**The role of cholesterol efflux in regulating the fertilization potential of mammalian spermatozoa. *J Clin Invest.* **110** (6):731-736.

**Werthman, P., R. Wixon, K. Kasperson and D. P. Evenson, (2007).**Significant decrease in sperm deoxyribonucleic acid fragmentation after varicocelelectomy. *Fertil Steril.* Published online June 11, White rabbit does. *Egyptian J. RabbitSci.*, 8(2): 157-167.

**Wilke, W., L. Wilke and O. Rowen, (2009):** Frandson and Anna Dec Fails. *Anatomy and Physiology of Farm animals.*, 746-757.

**Zalata, A. A., A. H. Ahmed, S. S. Allamaneni, F. H. Comhaire and A. Agarwal, (2004).**Relationship between acrosin activity of human spermatozoa and oxidative stress. *Asian J Androl*, 6:313–318.